

Induction of severe acute organophosphorus poisoning at the muscle level: what could we learn from zebrafish?

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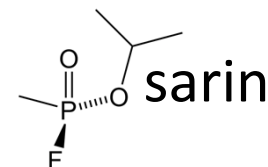
Financial support: NATO Science for Peace and Security Programme (MD.SFPP 984777)

International collaborative work

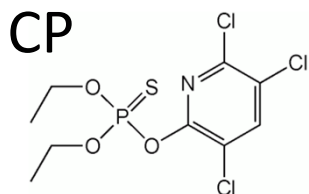
- IDAEA-CSIC, Barcelona, Spain (***Raldúa Demetrio and Prats Eva***)
- CESAM, University of Aveiro, Portugal (***Faria Melissa***)
- Universitat Autònoma de Barcelona, Spain (***Padrós Francesc***)
- Mississippi State University, USA (***Garcia-Reyero Natalia***)
- US Army Engineer Research and Development Center, Vicksburg, USA (***Arick II Mark***)
- CIB-CSIC, Madrid, Spain (***Rial Eduardo***)
- Universitat de Barcelona, Spain (***Sebastián David and Zorzano Antonio***)
- Université de Bordeaux, France (***Knoll-Gellida Anja and Babin Patrick J***)

Neuropathic organophosphorus (OPs) are used as:

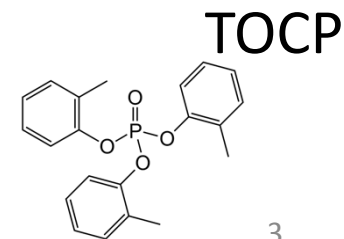
- **Chemical warfare agents** (*e.g.* sarin, soman)



- Pesticides in agriculture (*e.g.* chlorpyrifos)
- Plasticizer and other applications in industry (*e.g.* tri-o-cresyl phosphate)



“Toxic Industrial Chemicals” (TICs)



1) Neuropathic OPs used as **chemical warfare agents**:

- *Military and terrorist threat:*

(**Sarin** : Syria civil war (2013);
terrorist attack in Tokyo subway,
1995)



2) **Toxic Industrial chemicals (TICs)**:

OP pesticides = approximately 3
millions cases of acute severe poisoning
and 300,000 deaths annually.

- *Accidental exposure*

- *Suicide*

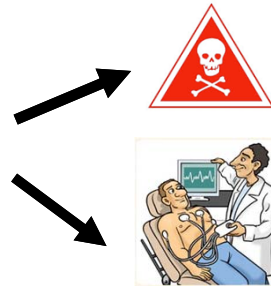


Neuropathic OPs are potent inhibitors of:

1. Acetylcholinesterase → Short term toxicity



Cholinergic syndrome



2. Neuropathic Target Esterase → Delayed toxicity



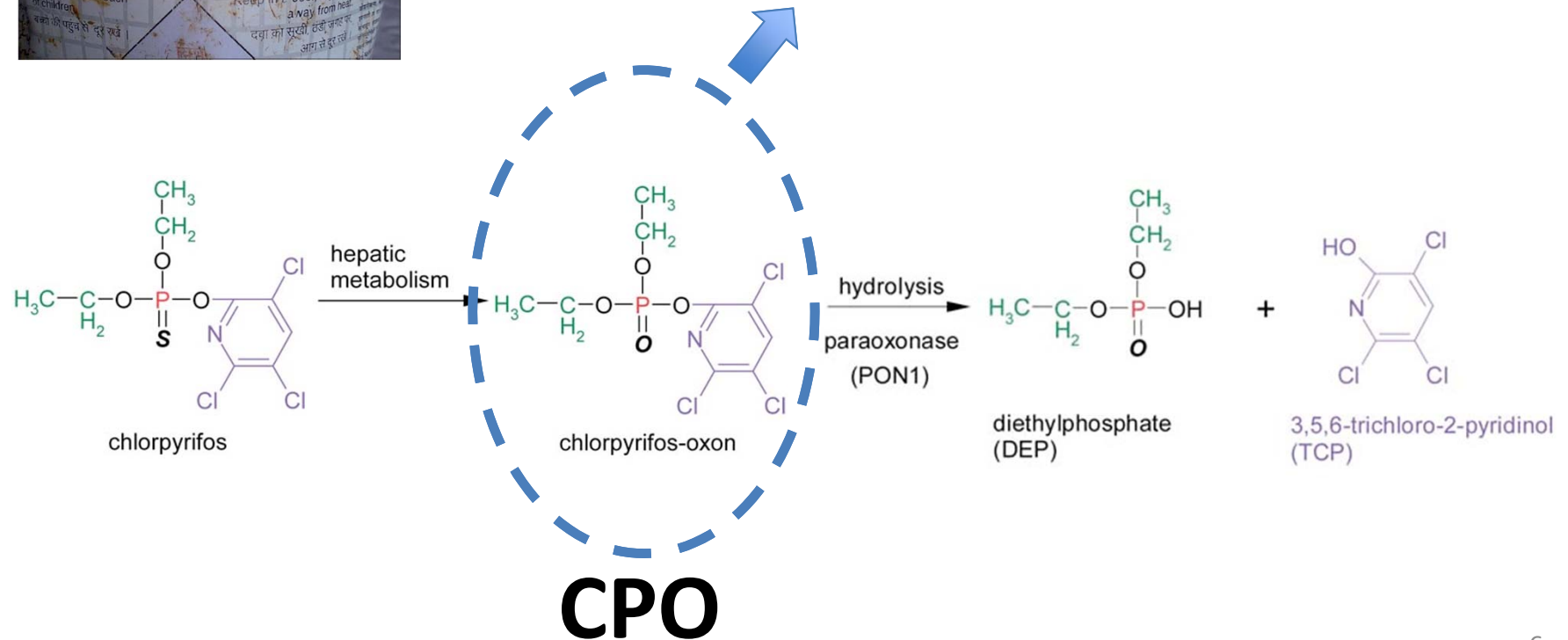
**Organophosphorus induced delayed neuropathy
(OPIDN)**



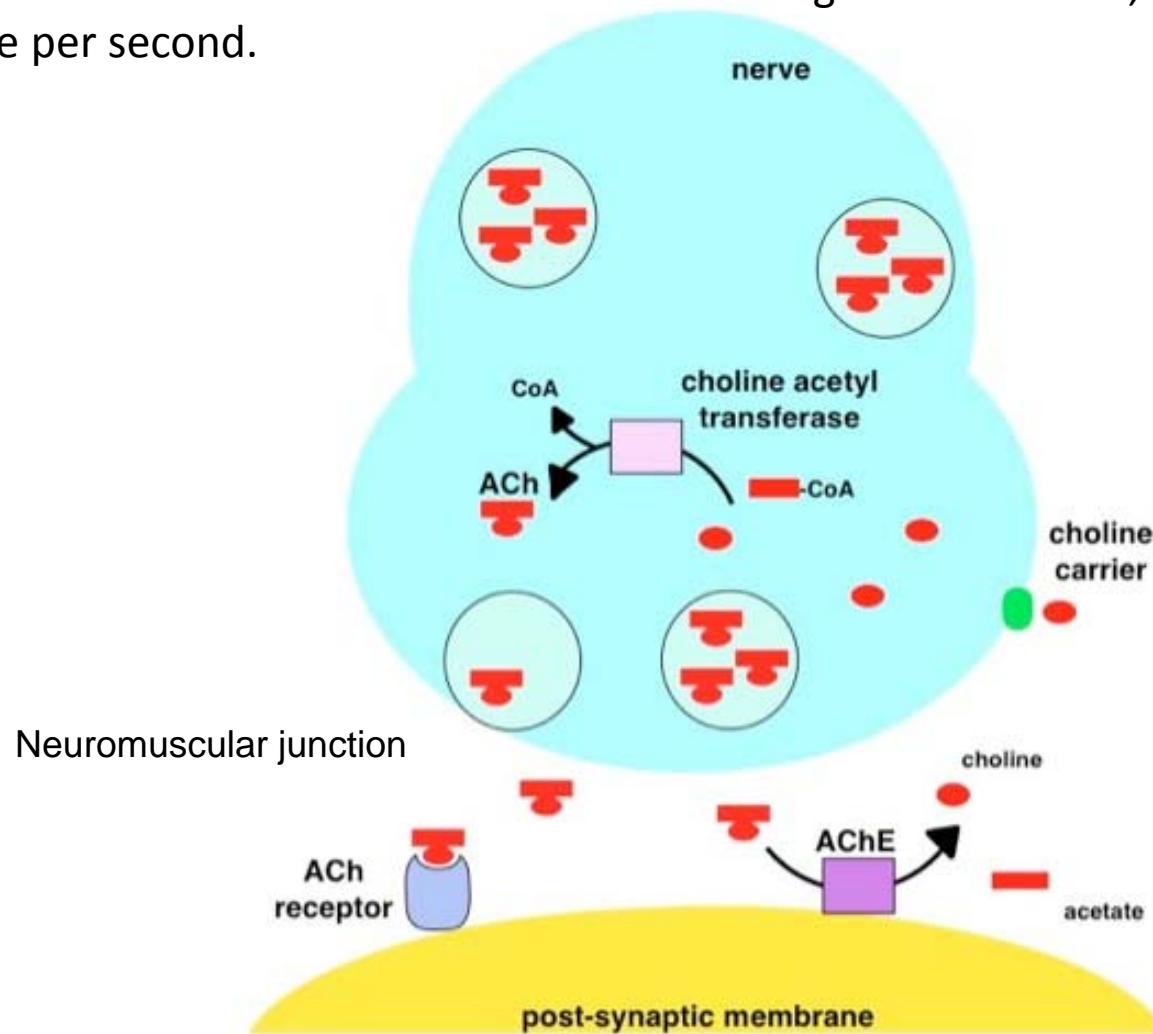
CHOLINERGIC TOXIDROME



ACETYLCHOLINESTERASE INHIBITION



Mechanism of action of AChE. Each AChE molecule degrades about 25,000 molecules of acetylcholine per second.



Manifestations of Organophosphate Poisoning

Optic System

Pupil Constriction
Blurred Vision
Lacrimation

Respiratory System

Bronchospasm
Bronchial Secretion
Pulmonary Edema
Tightness of Chest
Wheezing
Cough
Difficulty Breathing

Gastrointestinal Tract

Salivation
Nausea
Cramps
Abdominal Pain
Vomiting
Diarrhea
Fecal Incontinence

Urinary - Genital

Urinary Incontinence
Impotence
Uterus Contraction

Brain

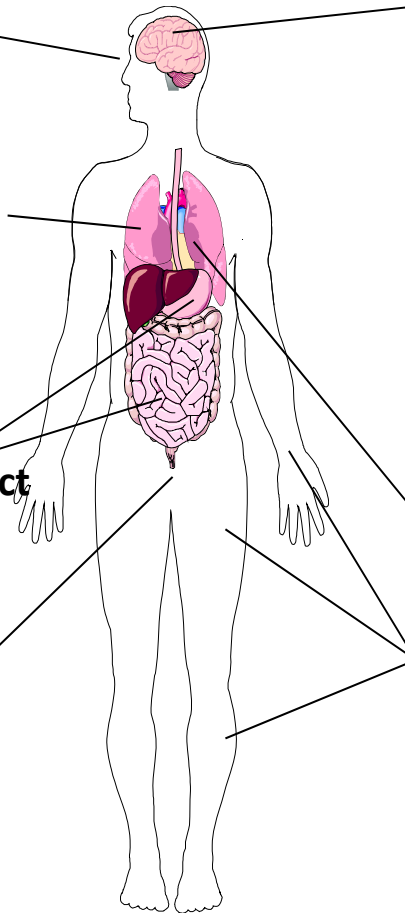
Headache
Dizziness
Vertigo
Anxiety
Apathy
Confusion
Anorexia
Insomnia
Lethargy
Fatigue
Inability to Concentrate
Memory Impairment
Convulsion
Coma

Cardiovascular System

Tachycardia
Increased Blood Pressure

Musculature

Weakness
Tremor
Fasciculations
Twitching
Cramps
Increased Sweating



General aim of the project

Zebrafish model(s) of cholinergic toxidrome



***In vivo* high-throughput screenings of small molecule libraries with zebrafish larva for identifying new drugs for multifunctional drug therapy against acute organophosphorus poisoning**

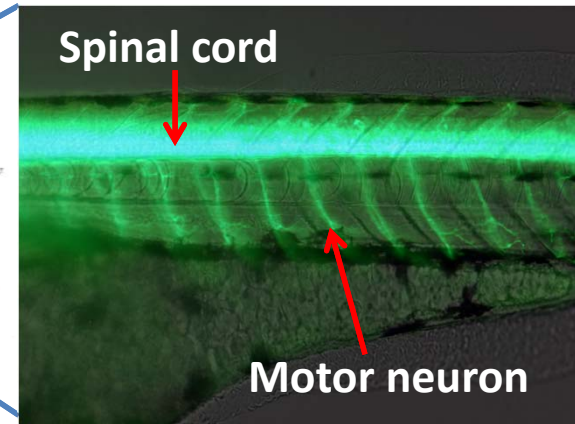
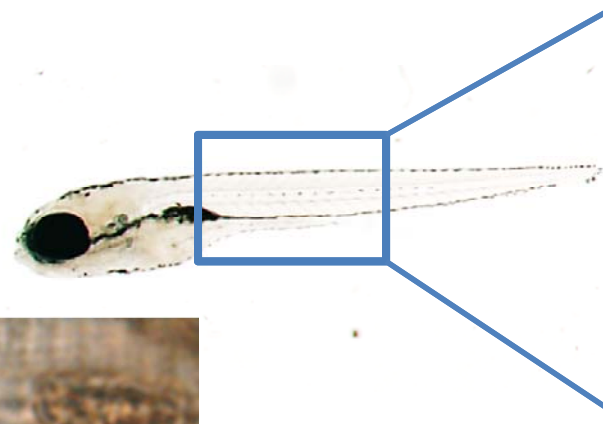
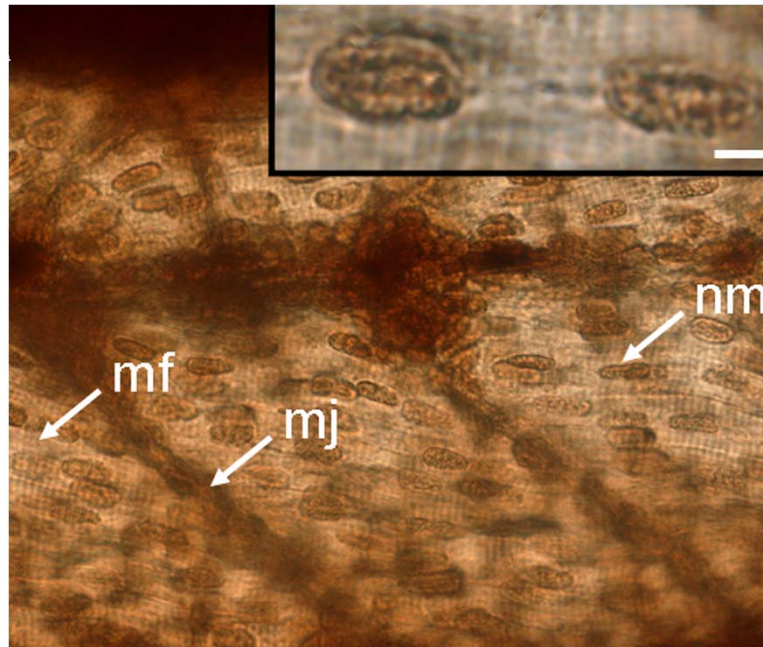
Zebrafish model of cholinergic toxidrome



- ✓ Inherent advantages: low cost, short life cycle, transparency at the larval stages, high number of larvae to be used in microplates, conservation with human of the biological functions investigated,

Zebrafish model of cholinergic toxidrome

- ✓ Genetic and cellular tools

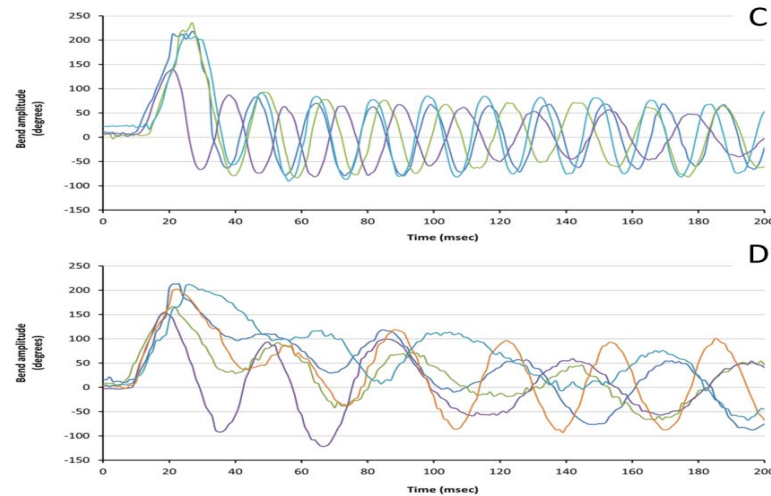
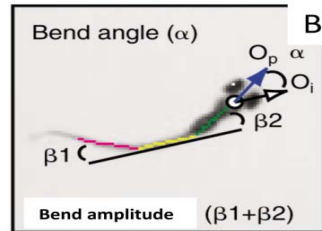
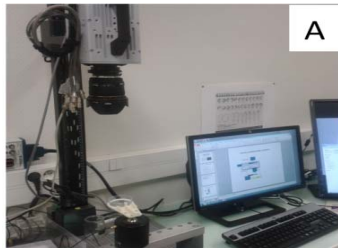


Live larva (e.g. transgenic line *hb9*)

Neuromuscular junction pattern and muscular fiber organization

Zebrafish model of cholinergic toxidrome

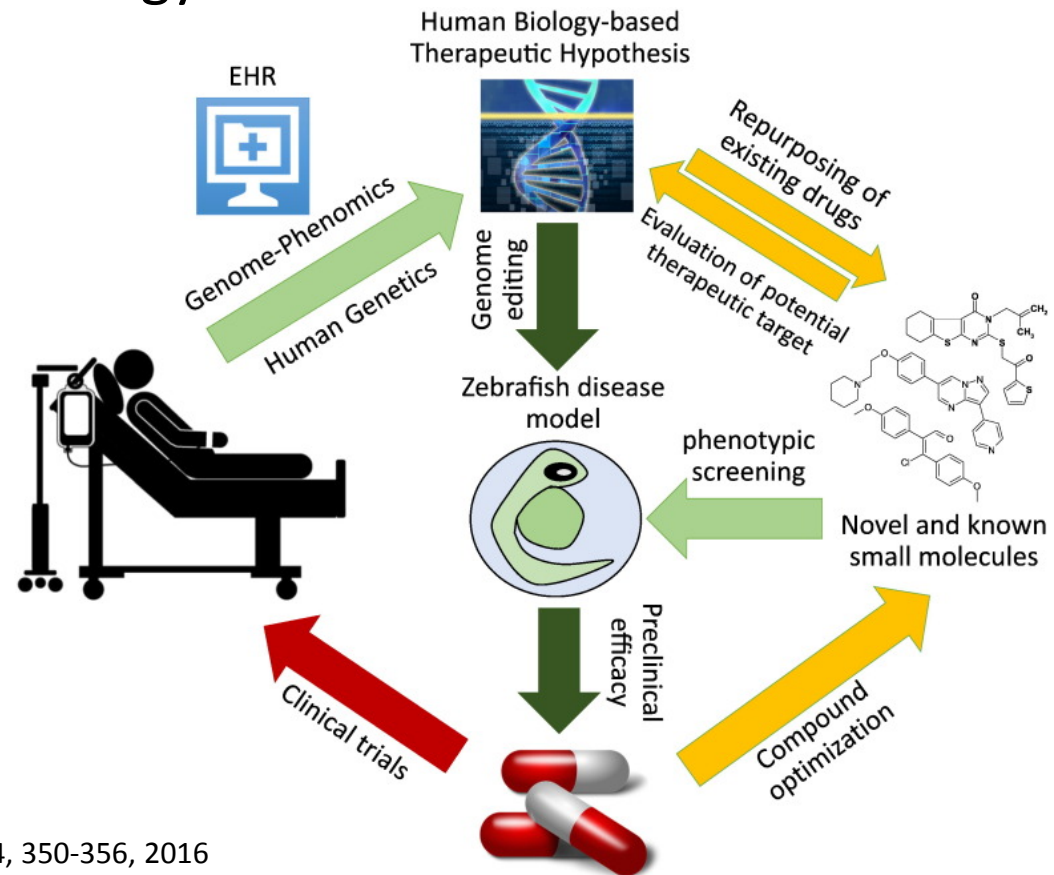
✓ Locomotor function assessment



Acoustic/Electric/Touch
escape response: kinematic
analysis

Zebrafish model of cholinergic toxidrome

✓ Pharmacology and toxicology

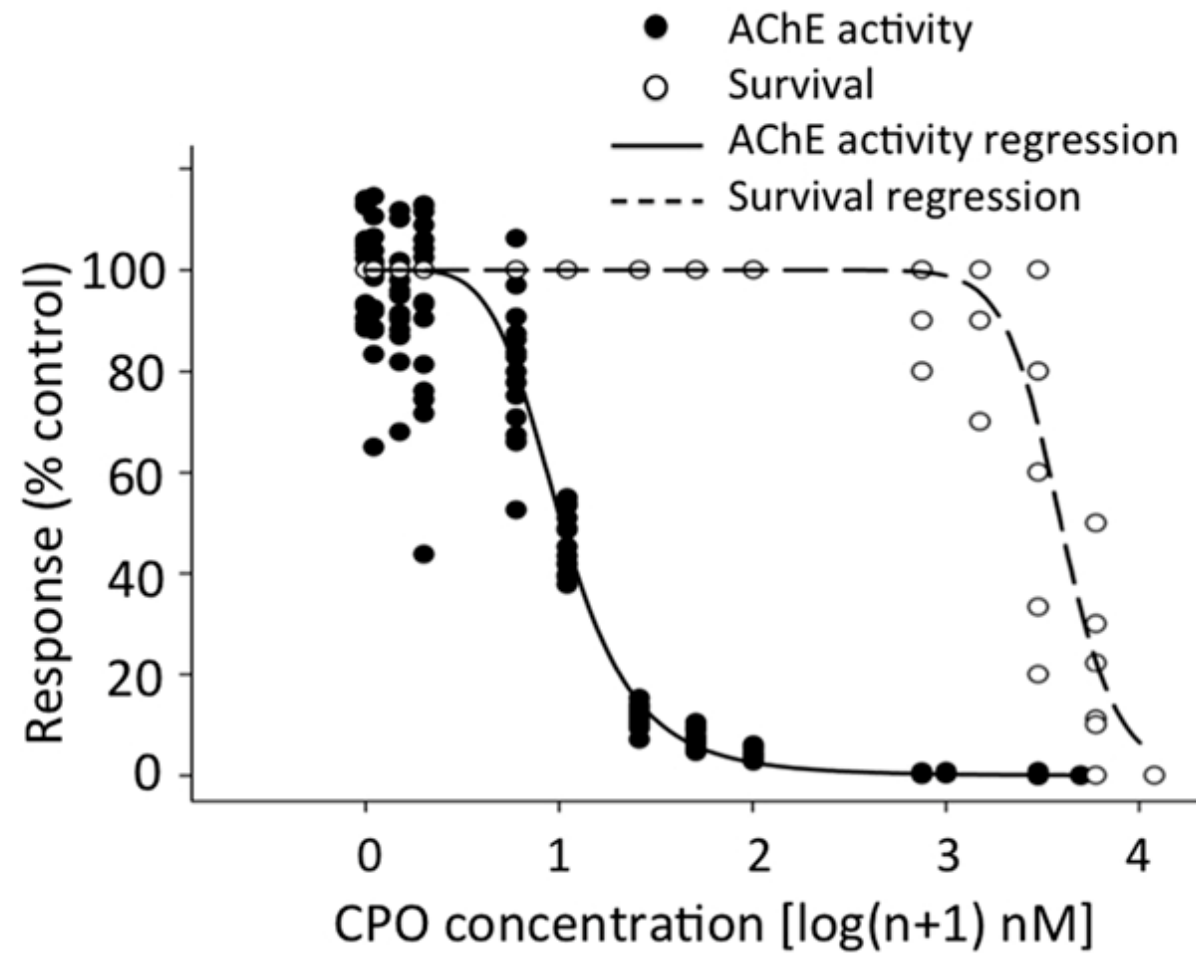


Zebrafish model of cholinergic toxidrome

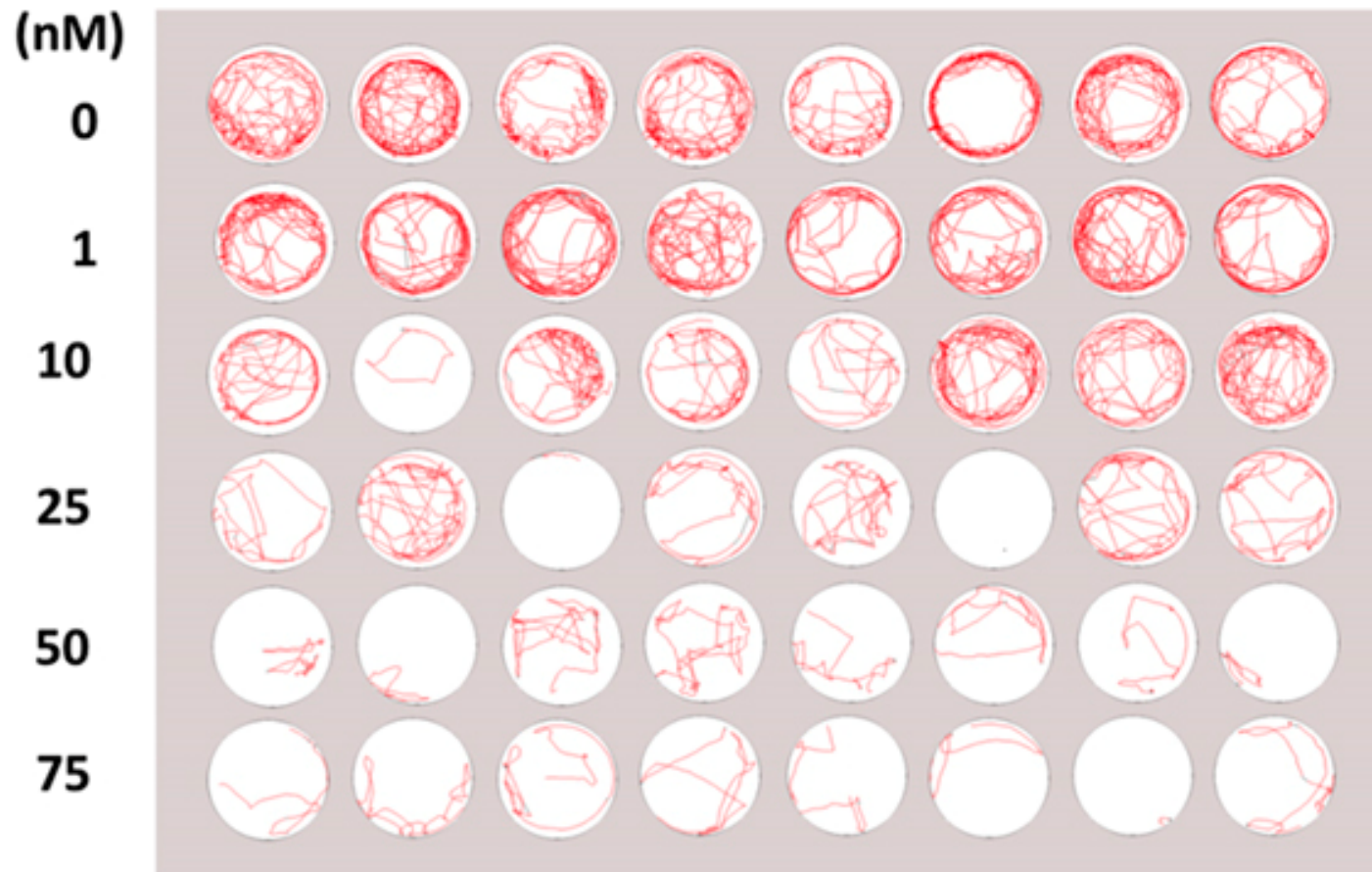


Generation and validation of zebrafish models for mild, moderate and severe acute OP poisoning by exposing zebrafish larvae to different concentrations of the prototypic OP compound, chlorpyrifos-oxon (CPO)

CPO induces concentration-dependent inhibition of acetylcholinesterase (AChE)

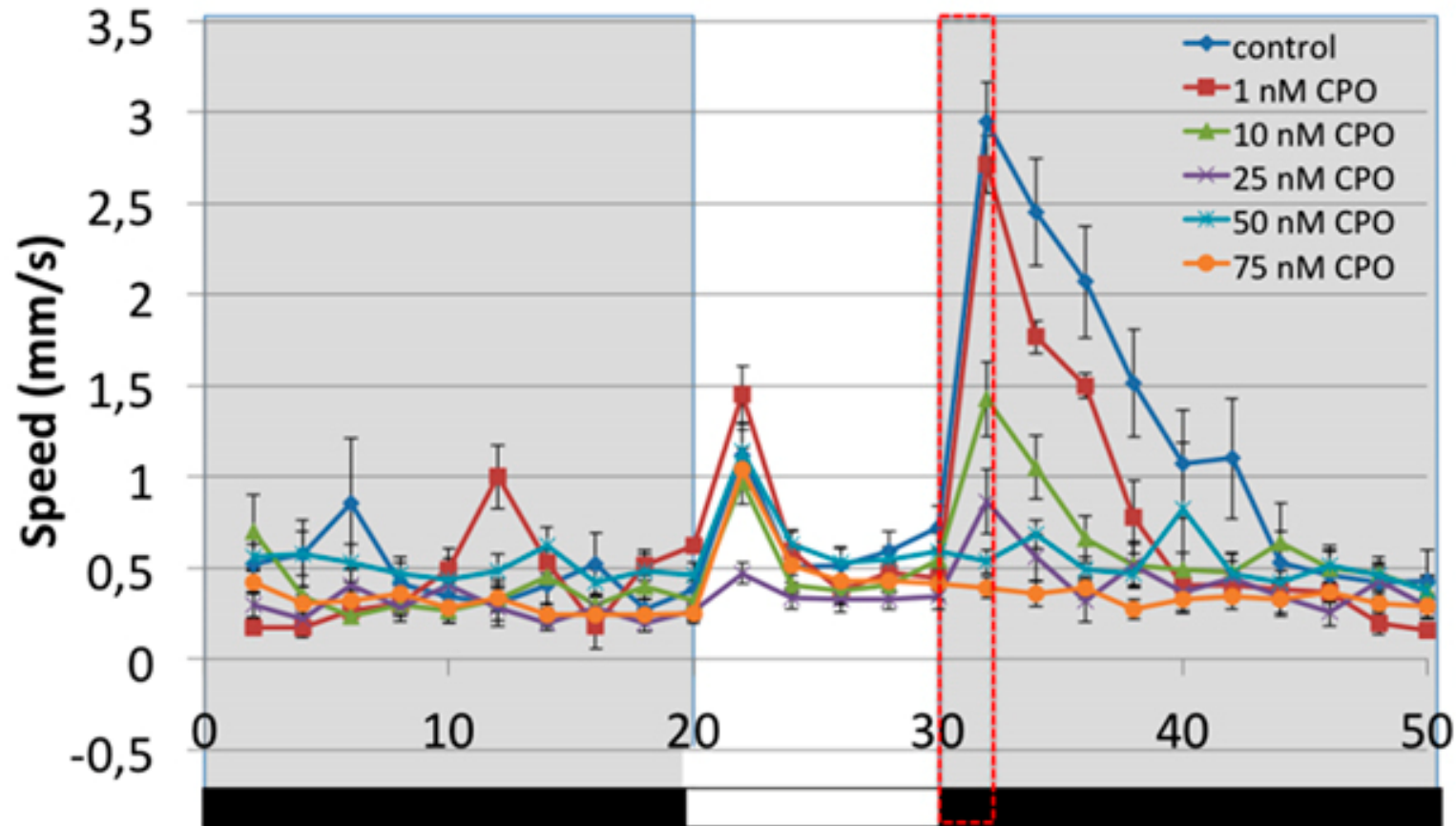


Motor behaviour is strongly impaired by CPO in zebrafish larvae



Locomotion tracking plots for the Visual Motor Response (VMR) assay in a 48-well behavioural arena ¹⁶

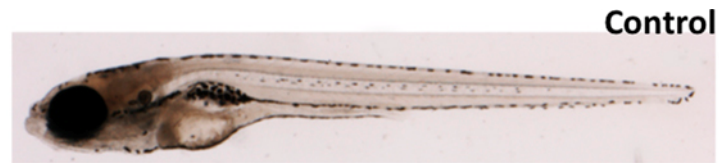
Motor behaviour is strongly impaired by CPO in zebrafish larvae



VMR profiles, showing that the hyperactivity peak evoked in response to sudden exposure to darkness is reduced until total abolition.

CPO induces the expression of three different phenotypes in zebrafish larvae according to CPO concentration

Day 8 larvae after
24 h exposure to CPO



Mild

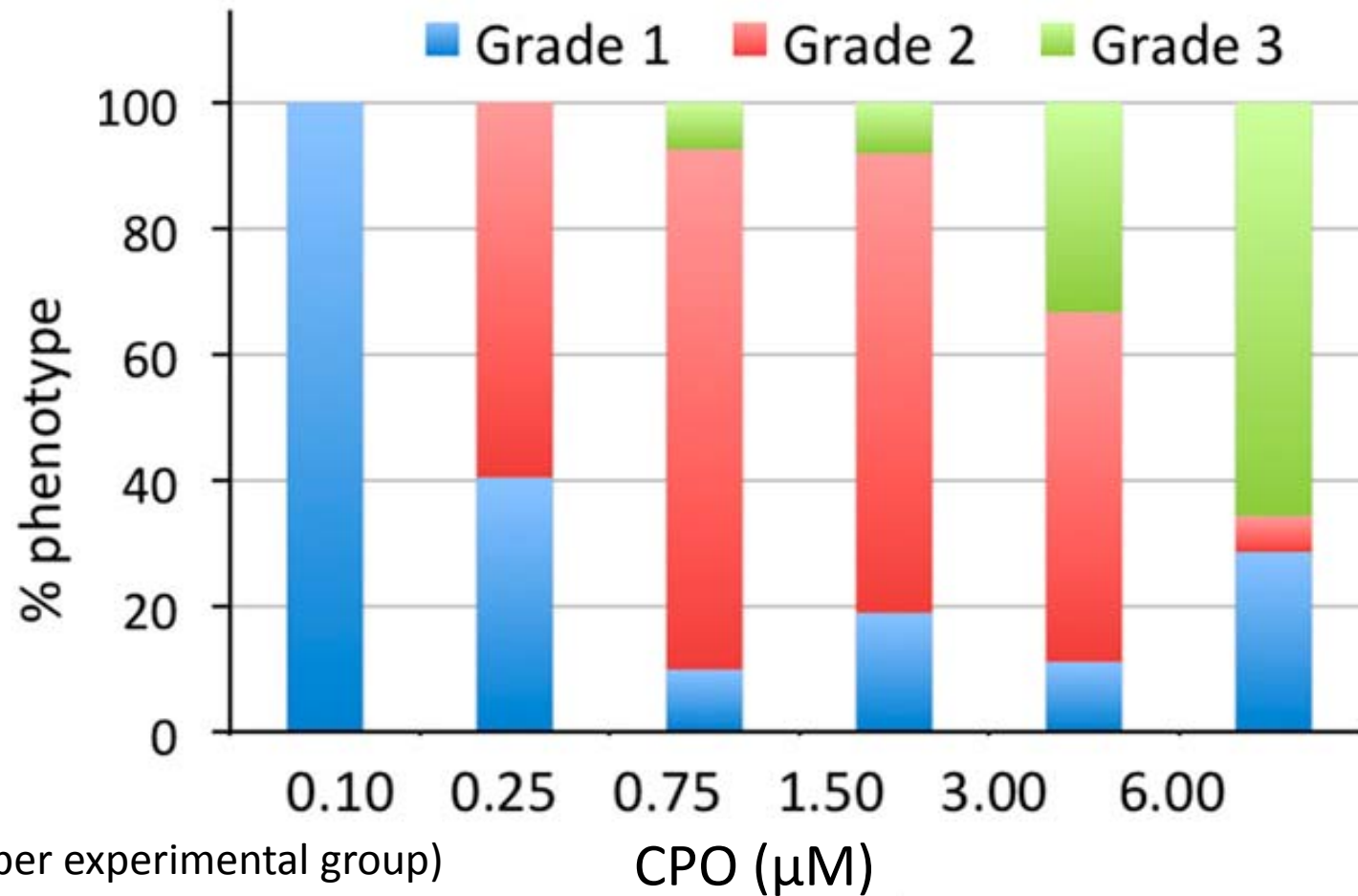


Moderate

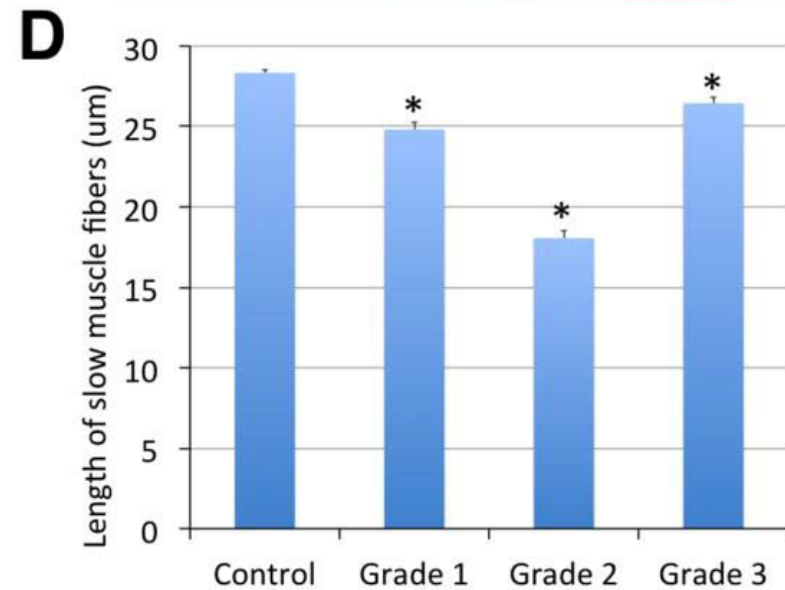
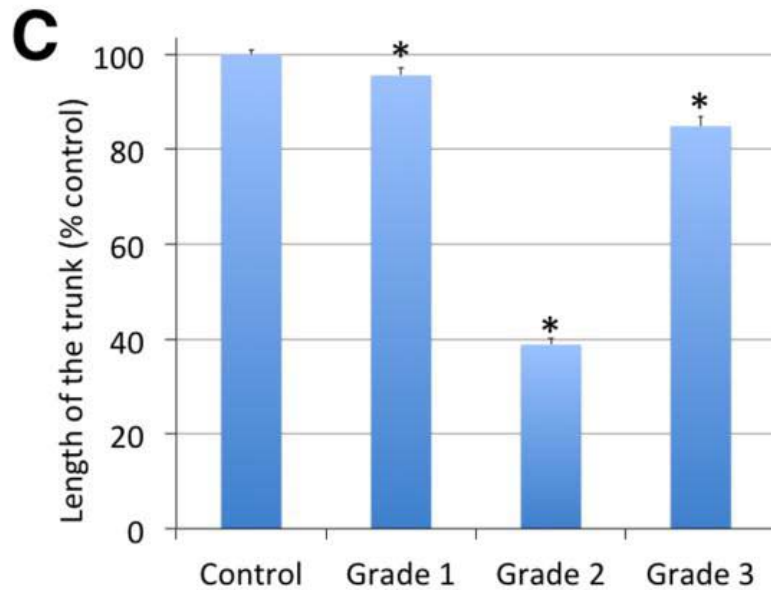
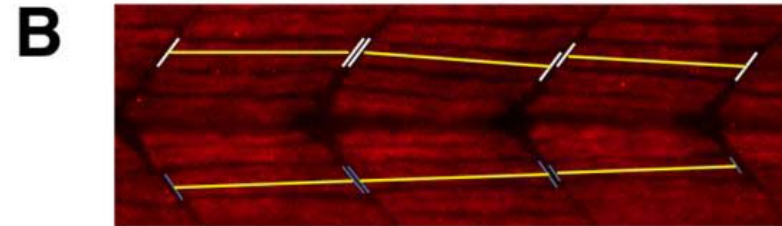
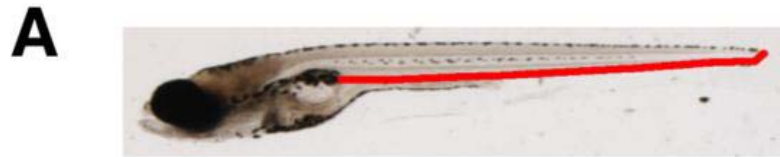


Severe

Prevalence of the three different phenotypes in response to different CPO concentrations




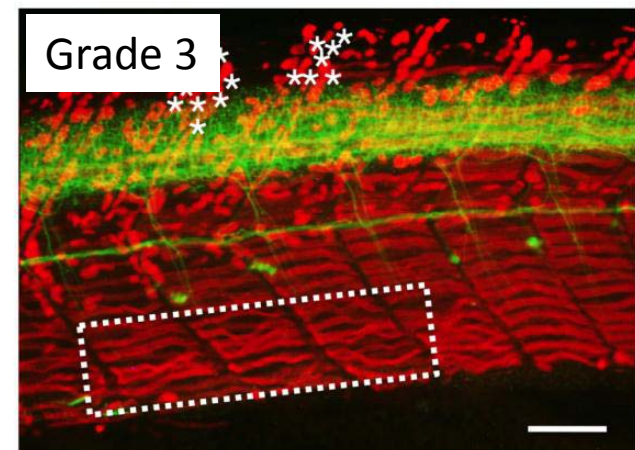
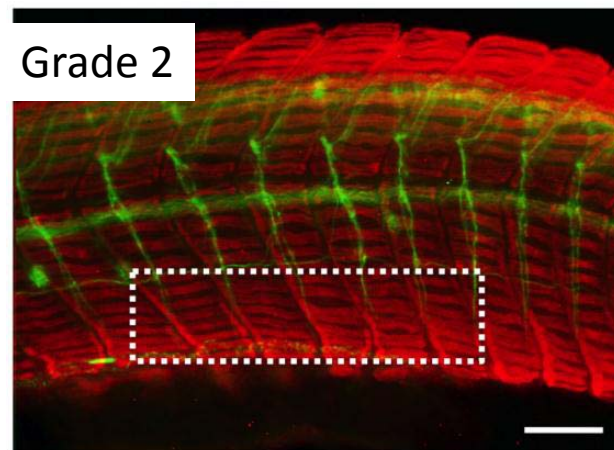
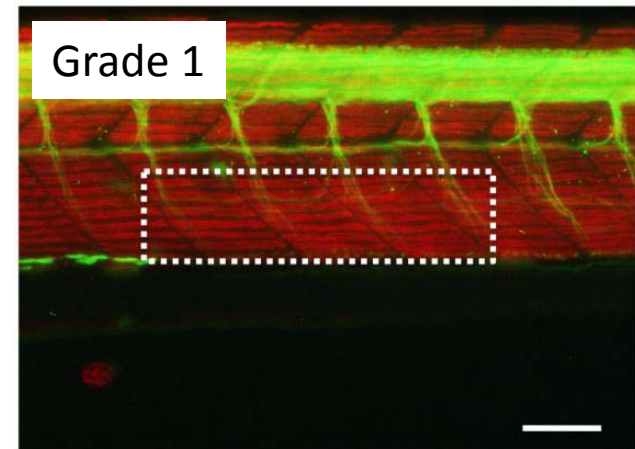
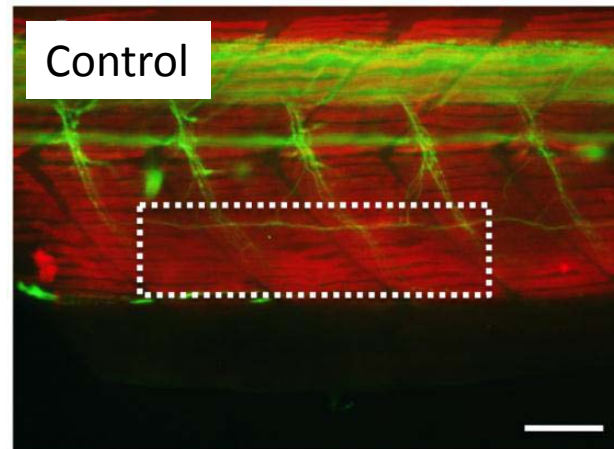
CPO induces reduction in the length of the trunk and axial muscle fibers in larvae



Characterization of pathological features at the trunk level in the different grades of severity of acute organophosphorus poisoning induced by chlorpyrifos-oxon in zebrafish larvae.

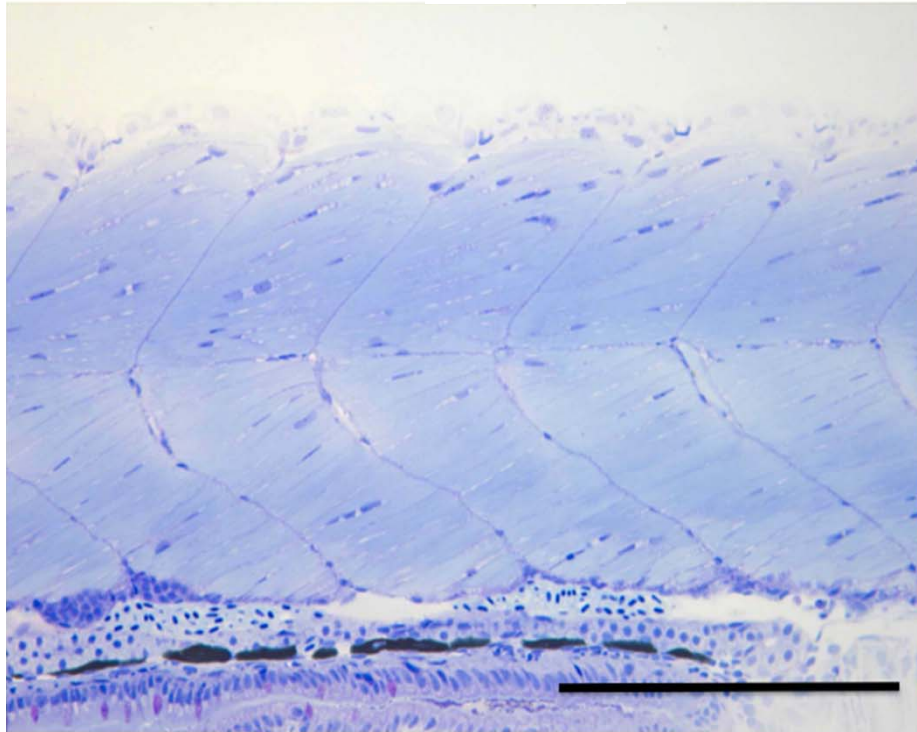
 F59 (slow muscle fibers)

 Acetylated α -tubulin (spinal axonal tracks)

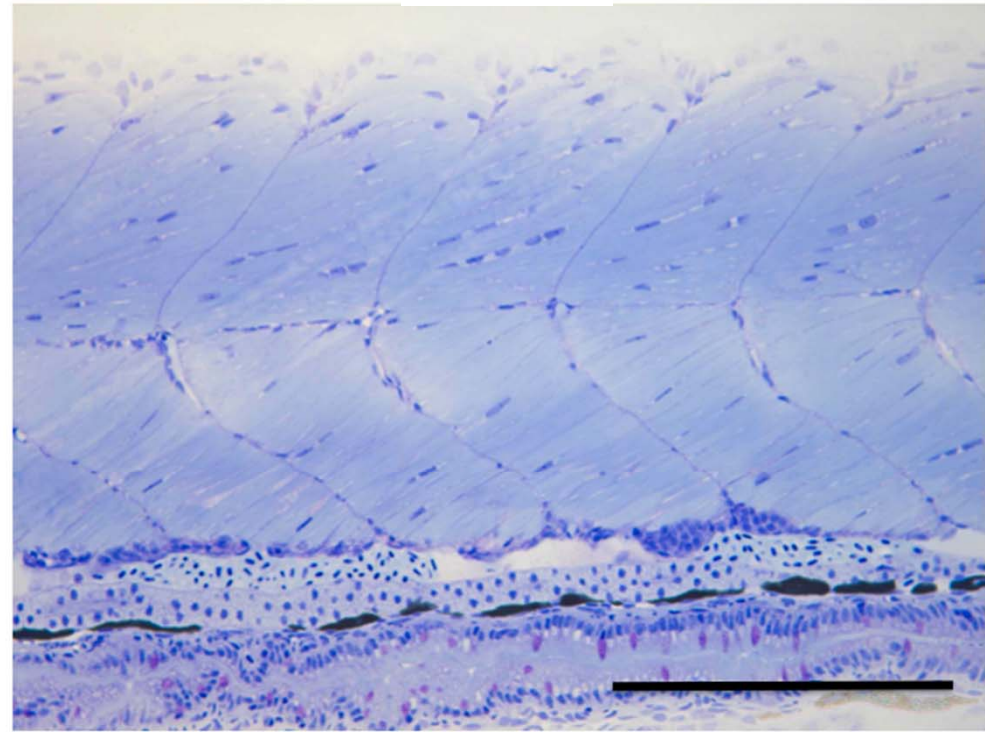


Histopathological analysis of **grade 1** phenotype at the muscle level

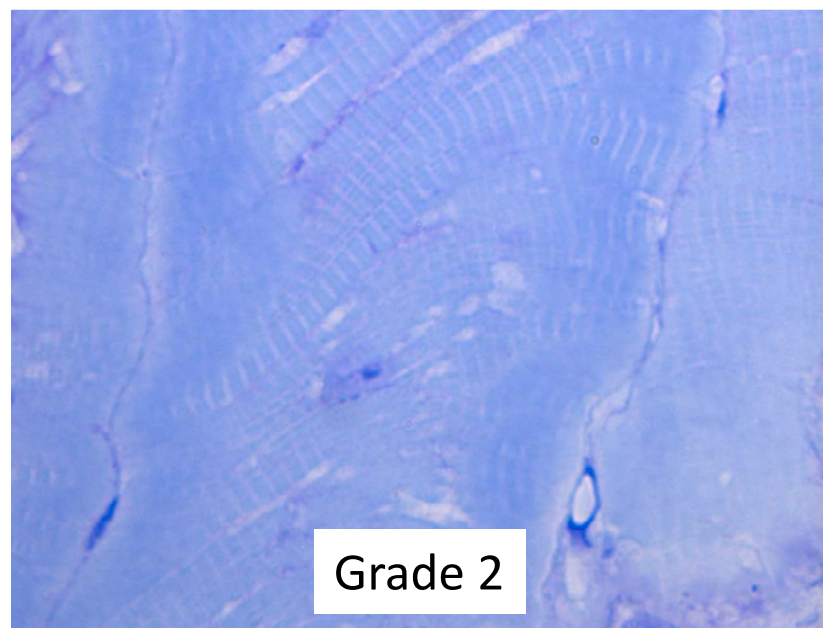
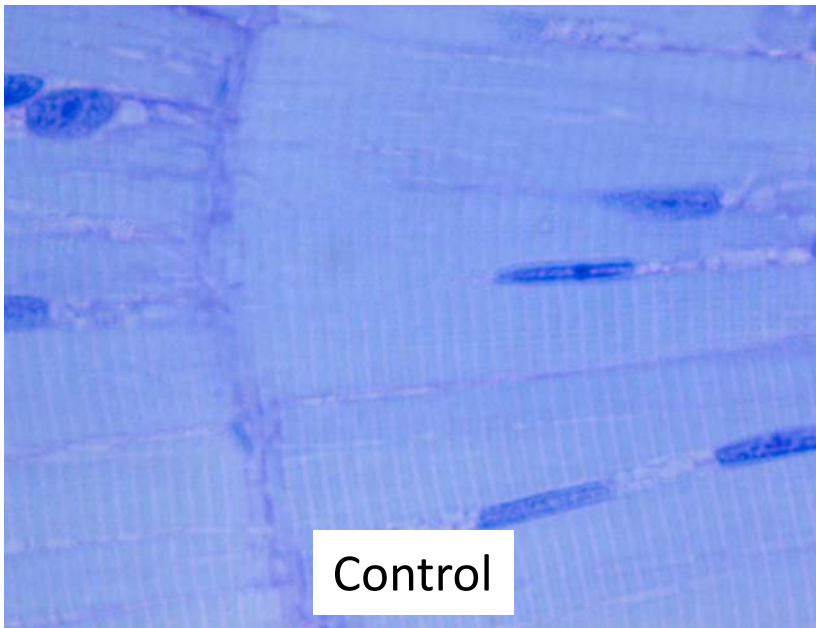
Control



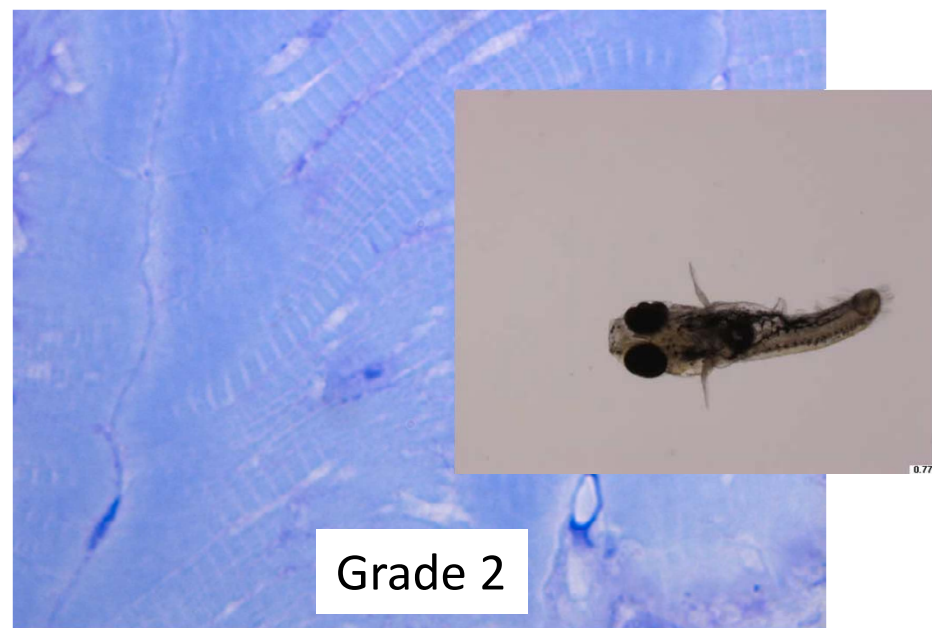
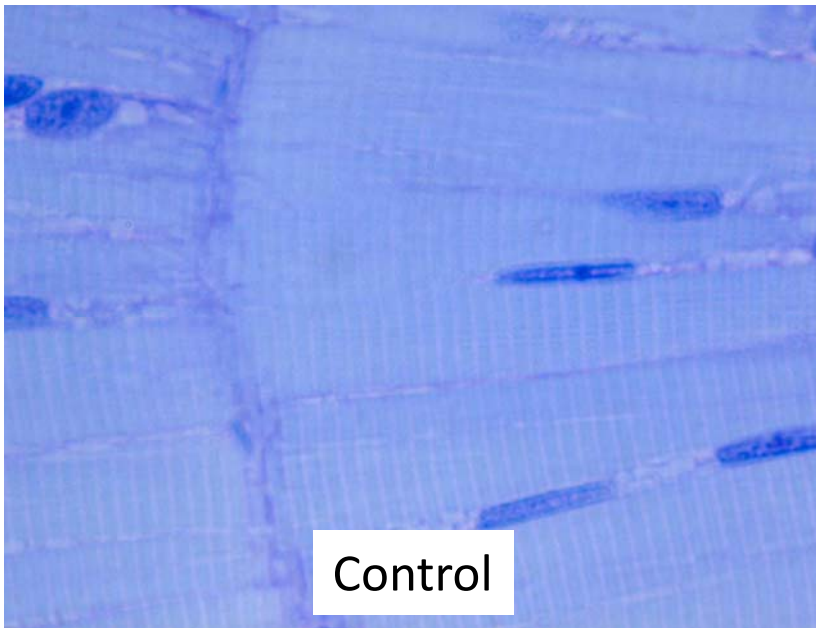
Grade 1



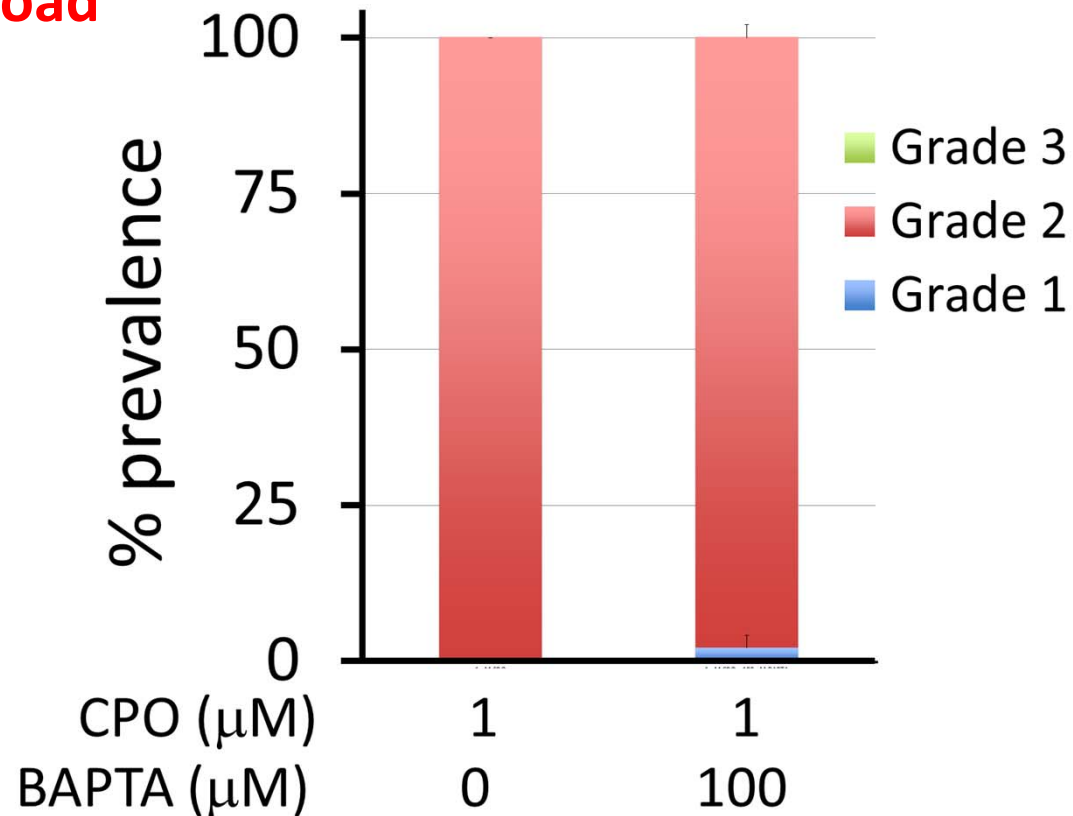
Hypercontracture of the axial muscles in **grade 2** larvae



Hypercontracture of the axial muscles in **grade 2** larvae

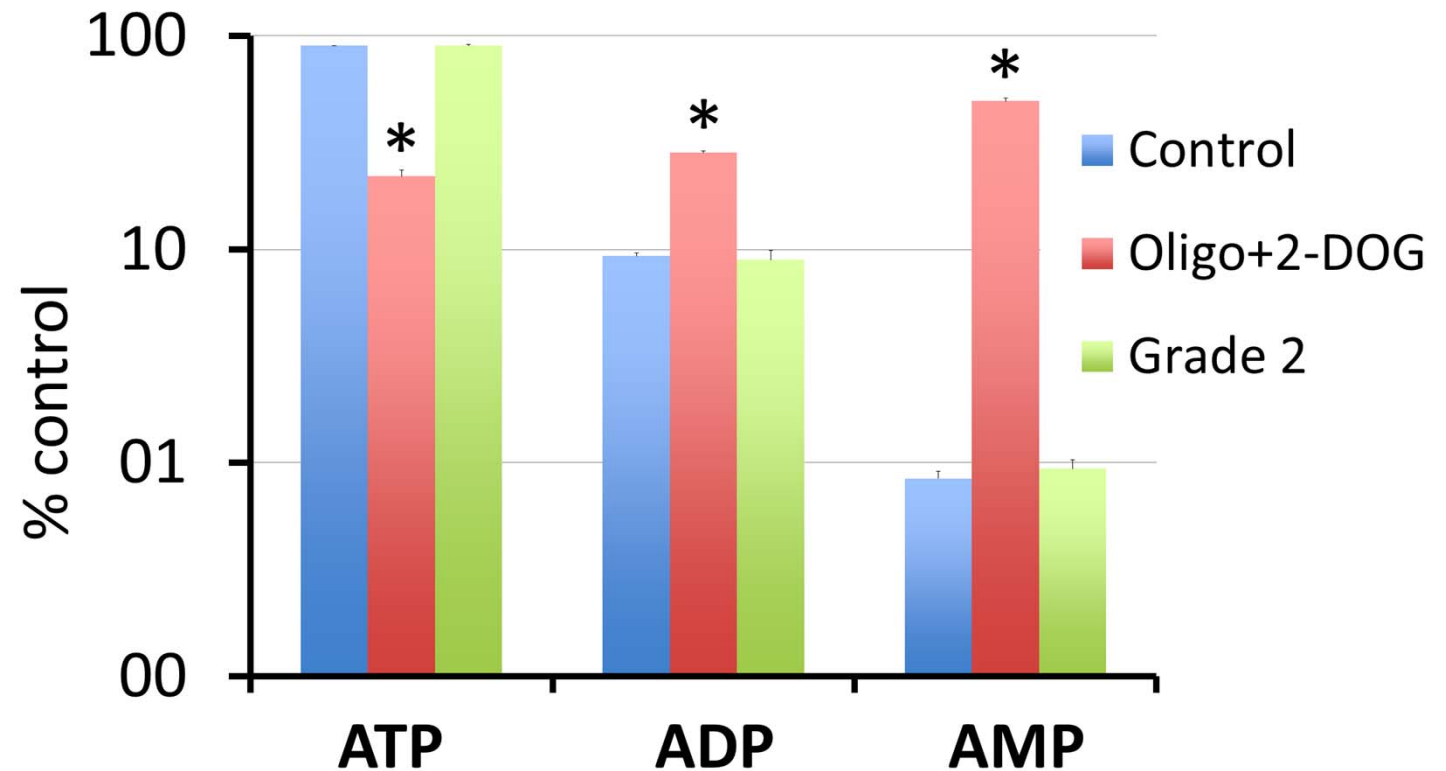


Hypercontracture of the axial muscles in grade 2 larvae **is not related with calcium overload**



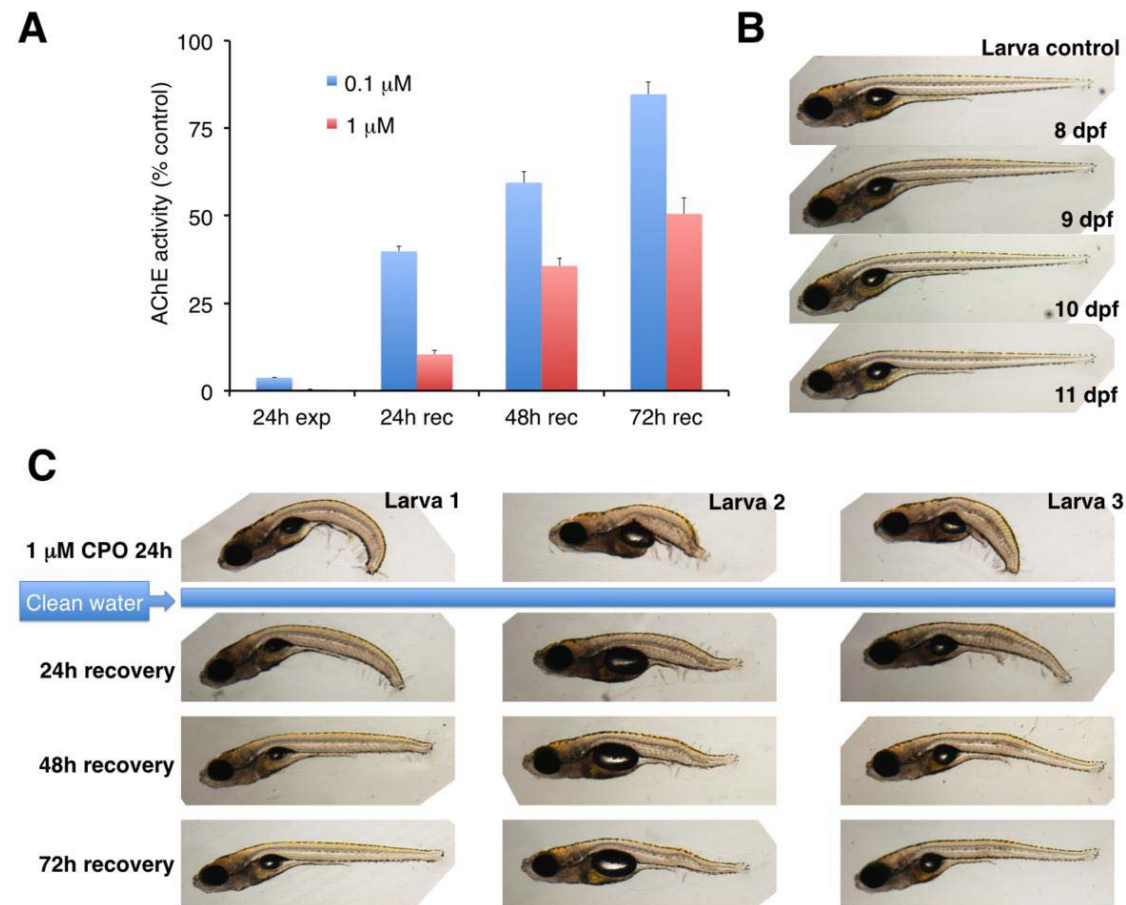
BAPTA-AM, a permeable calcium chelator, is not able to rescue the grade 2 phenotype, indicating that Ca^{2+} overload is not the mechanism leading to the hypercontracture.

Hypercontracture of the axial muscles in grade 2 larvae is not related with ATP depletion.



Oligomycin + 2-deoxyglucose (2-DOG)

Partial recovery of AChE activity and **grade 2** phenotype in zebrafish larvae exposed to 1 μ M CPO after the transfer to clean water.

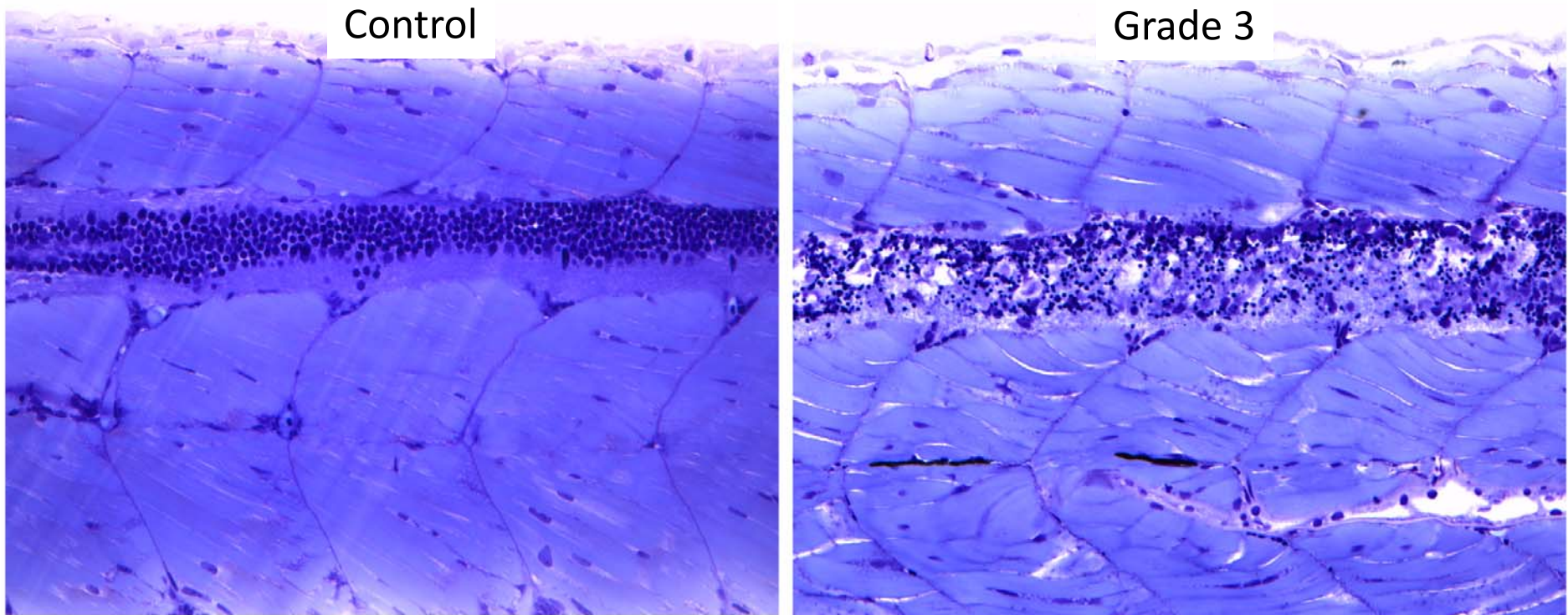


Grade 2 was the most prevalent phenotype within the 0.75–1.50 μM range of CPO concentrations. Larvae were unable to swim and their touch motor response was fully abolished.

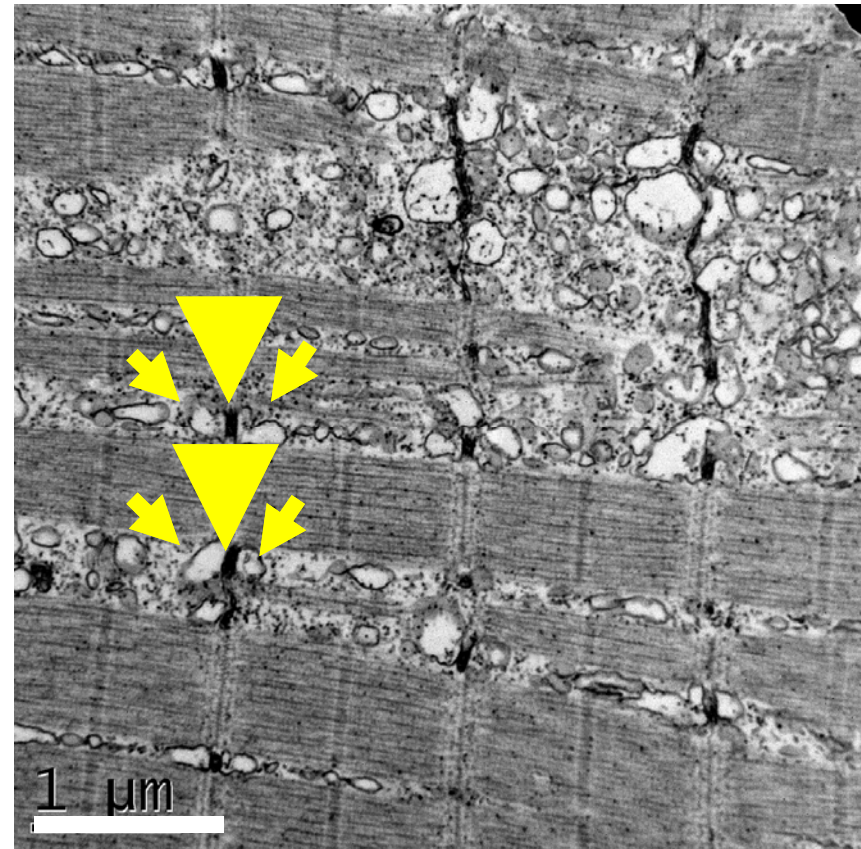
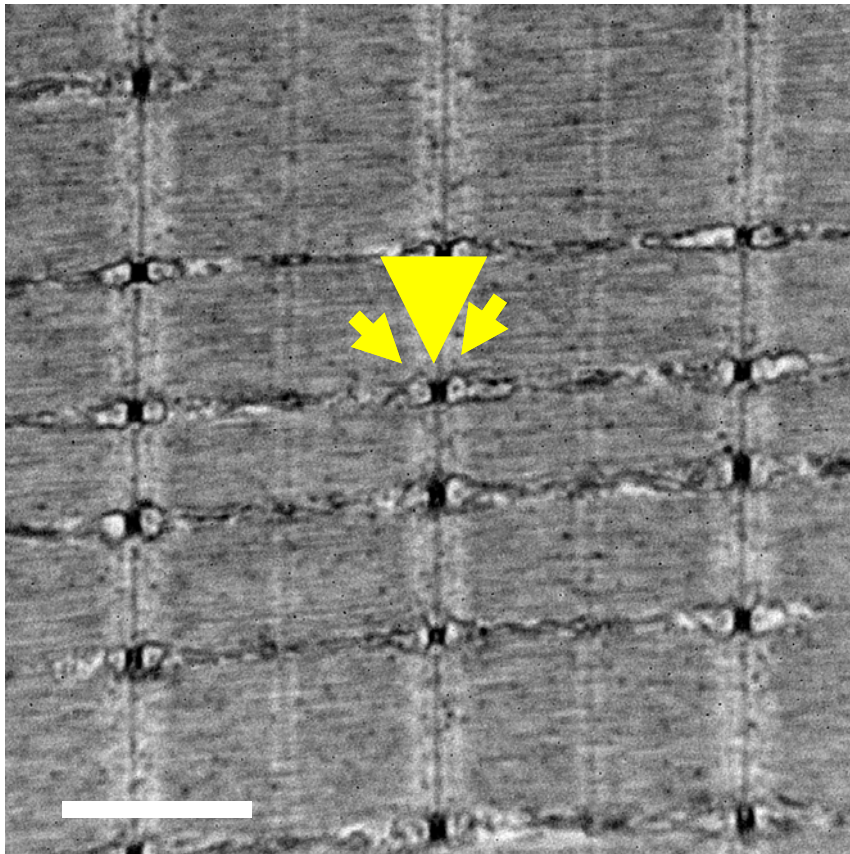
AChE inhibition was an upstream event in the toxic pathways that resulted in hypercontracture of the axial muscles and integrity impairment of axial muscle fibers which were not related with calcium overload or ATP depletion.

Reversibility, at least partial, of CPO's effects on body length and locomotor behavior may be observed after removing the compound from the water.

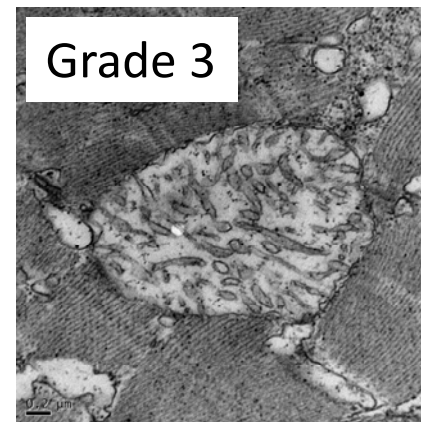
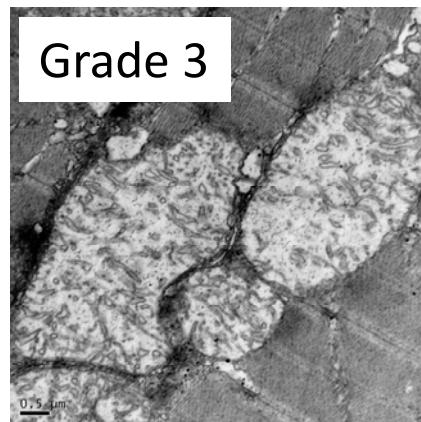
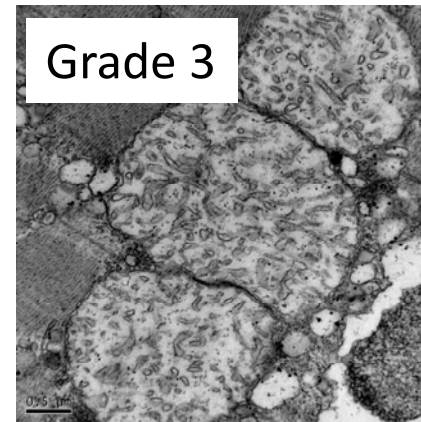
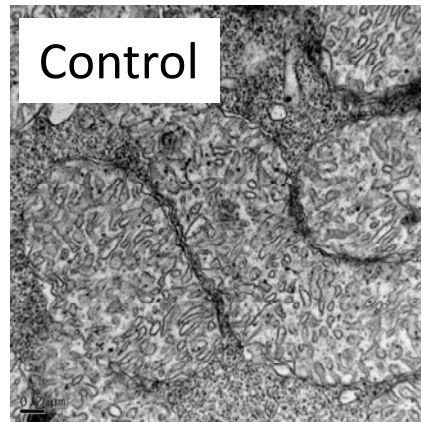
Histopathological assessment of **grade 3** larvae shows **severe lesions** in **muscle fibers**



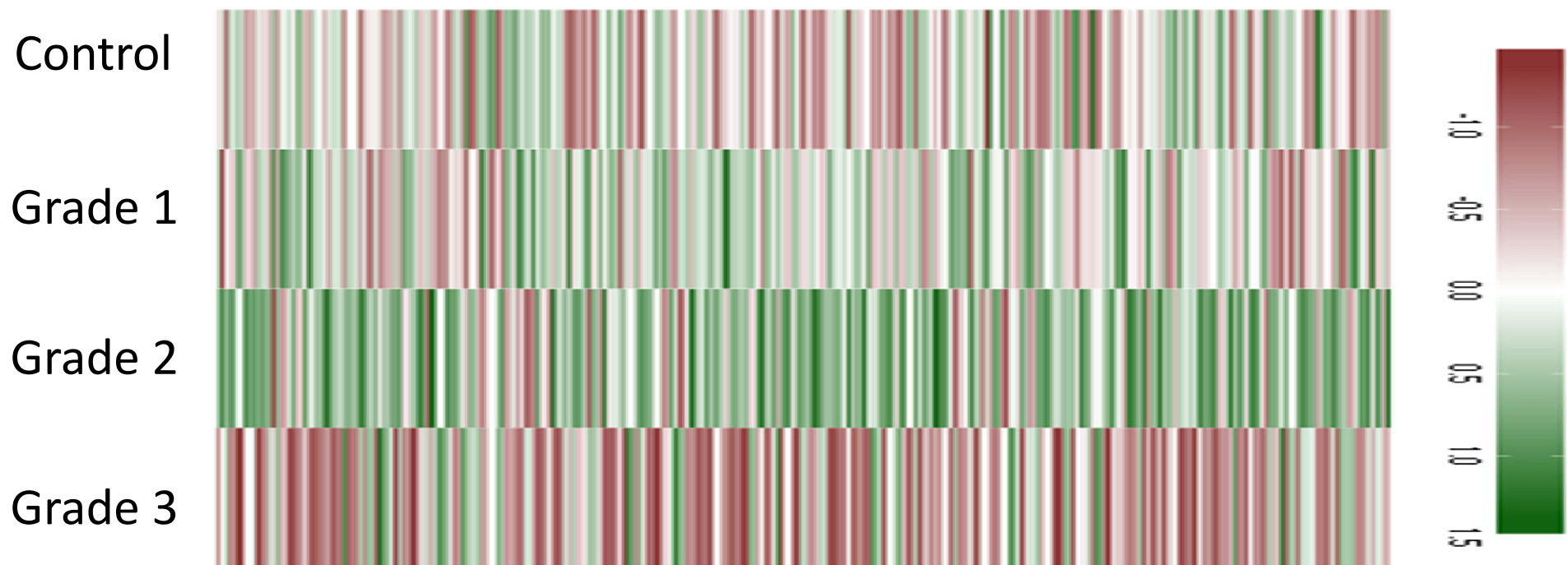
Histopathological assessment of **grade 3** larvae shows **severe lesions** in **muscle fibers**



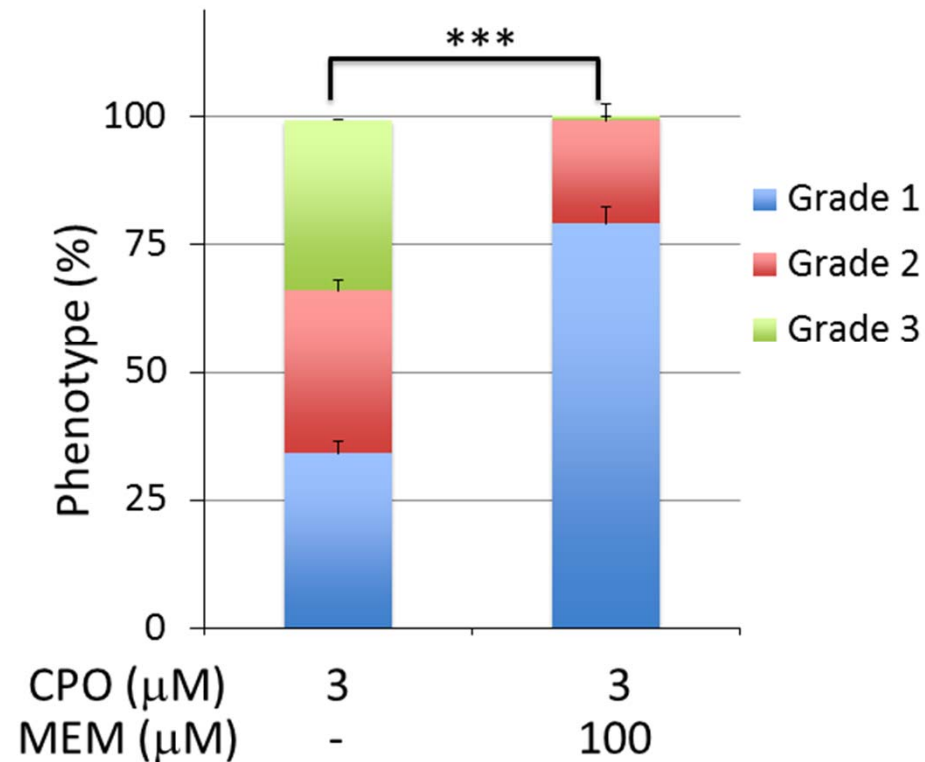
Swelling and loss of cristae in the mitochondria of the **grade 3** larvae (fast-twitch axial muscle fibers)



Calcium dysregulation is central to **grade 3** development. Heatmap of the **calcium signalling pathway** (dre04020) in control and the different grades of OPP, showing clear down-regulation of this pathway in grade 3 larvae.

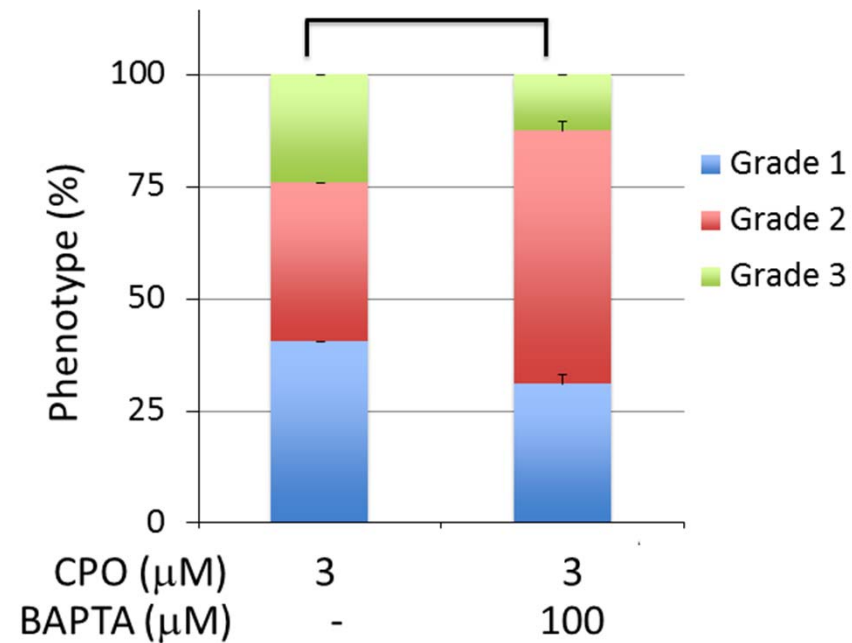


Calcium dysregulation is central to grade 3 development. Memantine, an antagonist of NMDA receptors, reduced the prevalence of the grade 3 phenotype dramatically.



Seven days post fertilization (dpf) larvae were pre-incubated with 100 μM memantine for 1 h followed by co-exposure with 100 μM memantine/3 μM CPO for 24 h.

Calcium dysregulation is central to grade 3 development. BAPTA-AM, a permeable calcium chelator, also reduced the prevalence of the grade 3 phenotype significantly.



Six days post fertilization zebrafish larvae were pre-incubated with 100 μM BAPTA-AM for 24 h followed by 24 h co-exposure with 3 μM CPO.

Grade 3 was the most prevalent phenotype at high CPO concentrations. It was characterized by an irreversible widespread necrosis of the axial muscle fibers. RNA-seq and pharmacological analyses supported the hypothesis that after initial cholinergic overstimulation, an influx of extracellular Ca^{2+} through NMDA receptors occurs and that the increase in the cytoplasmic levels of Ca^{2+} in animals with a compromised Ca^{2+} buffer capacity resulted in the uncontrolled activation of proteases, phospholipases and kinases.

General conclusion:

Our results show that zebrafish models may mimic pathophysiological mechanisms behind cholinergic toxidrome in humans. The suitability of the zebrafish larvae to *in vivo* high-throughput screenings of small molecule libraries makes these models a valuable tool for identifying new drugs for multifunctional drug therapy against acute organophosphorus poisoning including deleterious effect at the muscle level.